

MICROBIAL α -AMYLASES AND THEIR INDUSTRIAL APPLICATIONS: A REVIEW

DrKiroMojsov*

Abstract:

The biotechnological potential of α -amylases from microorganisms has drawn a great deal of attention from various researchers worldwide as likely biological catalysts in a variety of industrial processes. The rapid developments in the field of genetic engineering have given a new impetus to the biotechnology. Biotechnology also offers the potential for new industrial processes that require less energy and are based on renewable raw materials and environmentally healthy practices. This work represents a review of α -amylase family and the major characteristics, microbial sources, production, properties, industrial applications as highly demanded industrial enzyme in various sectors such as food, textiles, detergents, pharmaceuticals, etc. The review intends to explore the potential of these enzymes and to encourage new α -amylase-based industrial technology.

Key words: microbial α -amylase, enzyme characteristics, production, industrial applications, starch.

*Assistant Professor, Dept. of Textil Technology, University "Goce Delcev" Stip, Macedonia.

1.Introduction:

Enzymes are globular proteins and like other proteins consist of long chains of amino acids that fold to produce a three-dimensional product. Each unique amino acid sequence produces a specific structure, which has unique properties. Enzymes are responsible for many essential biochemical reactions in microorganisms, plants, animals, and human beings. They differ in function in that they have the unique ability to facilitate biochemical reactions without undergoing change themselves. This catalytic capability is what makes enzymes unique.

Enzymes, biological catalysts with high selectivities, have been used in the food industry for hundreds of years, and play an important role in many other industries (washing agents, textile manufacturing, pharmaceuticals, pulp and paper). They without being consumed in the process, can speed up chemical processes that would otherwise run very slowly, or in some cases, not at all [Cavaco-Paulo & Gübitz 2003]. After the reaction is complete, the enzyme is released again, ready to start another reaction. Usually most enzymes are used only once and discarded after their catalytic action. All known enzymes are proteins. They therefore consist of one or more polypeptide chains and display properties that are typical of proteins. Some enzymes require small non-protein molecules, known as cofactors, in order to function as catalysts [Jenkins 2003].

Currently, enzymes are becoming increasingly important in sustainable technology and green chemistry. Generally they are active at mild temperatures. Above certain temperature the enzyme is denatured. Enzymes have a characteristic pH at which their activity is maximal. Extreme pH values influence on the electrostatic interactions within the enzyme, leading to inactivation of enzyme. Other important factors that influence the effect of enzymatic processes are the concentration of enzyme, the time of treatment, additives like surfactants and chelators and mechanical stress [Tavčer 2011]. Enzyme can break down particular compounds. The molecule that an enzyme acts on is known as its substrate, which is converted into a product or products. Some of the most common include amylases which break down starch into simple sugars, proteases which break down proteins, cellulases which break down cellulose, and lipases which split fats (lipids) into glycerol and fatty acids. For each type of reaction in a cell there is a different enzyme and they are classified into six broad categories namely hydrolytic, oxidising and reducing, synthesising, transferring, lytic and isomerising. The essential characteristic of enzymes is catalytic function. Consequently, the original attempt to classify enzymes was done according

to function. The International Commission on Enzymes (EC) was established in 1956 by the International Union of Biochemistry (IUB), in consultation with the International Union of Pure and Applied Chemistry (IUPAC), to put some order to the hundreds of enzymes that had been discovered by that point and establish a standardized terminology that could be used to systematically name newly discovered enzymes. The EC classification system is divided into six categories of basic function:

- EC1 Oxidoreductases: catalyze oxidation/reduction reactions.
- EC2 Transferases: transfer a functional group.
- EC3 Hydrolases: catalyze the hydrolysis of various bonds.
- EC4 Lyases: cleave various bonds by means other than hydrolysis and oxidation.
- EC5 Isomerases: catalyze isomerization changes within a single molecule.
- EC6 Ligases: join two molecules with covalent bonds.

Each enzyme is described by a sequence of four numbers preceded by “EC”. The first number broadly classifies the enzyme based on its mechanism.

All α -amylases (EC3.2.1.1) are starch-degrading enzymes that catalyze the hydrolysis and act on internal α -1,4-glycosidic linkages in starch in low molecular weight products, such as glucose, maltose and maltotriose units [Gupta et al. 2003; Kandra 2003; Rajagopalan and Krishnan 2008]. Amylases are among the most important enzymes and are of great significance for biotechnology, constituting a class of industrial enzymes having approximately 25% of the world enzyme market [Rajagopalan and Krishnan 2008; Reddy et al. 2003]. Most of the α -amylases are metalloenzymes, which require calcium ions (Ca^{2+}) for their activity, structural integrity and stability. They belong to family 13 (GH-13) of the glycoside hydrolase group of enzymes [Bordbar et al. 2005].

The amylase family of enzymes is of great significance due to its wide area of potential application. Interestingly, the first enzyme produced industrially was an amylase from a fungal source in 1894, which was used as a pharmaceutical aid for the treatment of digestive disorders [Crueger W. and Crueger A. 1989]. Amylases find potential application in a number of industrial processes such as in the food, fermentation, textiles and paper industries. Microbial amylases have successfully replaced the chemical hydrolysis of starch in starch-processing industries. They would be potentially useful in the pharmaceutical and fine chemicals industries if enzymes with suitable

properties could be prepared [Fogarty and Kelly 1980]. The spectrum of amylase application has widened in many other fields, such as

clinical, medical, and analytical chemistries, as well as their wide spread application in starch saccharification and in the textile, food, fermentation, paper, brewing and distilling industries [Pandey et al. 2000].

Historically, the application of enzymes in industrial textile processes began around 1857, when malt extract was used to remove size from fabrics before printing [Ciechanska and Kazimierzak 2006; Marcher et al. 1993]. Starch is widely used as a sizing agent, being readily available, relatively cheap and based on natural, sustainable raw materials [Lange 1997]. About 75% of the sizing agents used worldwide are starch and its derivatives [Opwis et al. 1999]. There are several processes in the medicinal and clinical areas that involve the application of amylases [Sutton et al. 1999; Chiu and Chandler 1995; Becks et al. 1995; Menzel et al. 1998; Chelly et al. 1996; Strandberg et al. 1999]. Due to the increasing demand for these enzymes in various industries, there is enormous interest in developing enzymes with better properties such as raw starch degrading amylases suitable for industrial applications and their cost effective production techniques [Burhan et al. 2003].

They can be obtained from several sources, such as plants, animals and microorganisms. Today a large number of microbial amylases are available commercially and they have almost completely replaced chemical hydrolysis of starch in starch processing industry. The amylases of microorganisms have a broad spectrum of industrial applications as they are

more stable than when prepared with plant and animal α -amylases [Tanyildizi et al. 2005]. The major advantage of using microorganisms for the production of amylases is the economical bulk production capacity and the fact that microbes are easy to manipulate to obtain enzymes of desired characteristics.

α -Amylase has been derived from several fungi, yeasts and bacteria. However, enzymes from fungal and bacterial sources have dominated applications in industrial sectors [Gupta et al. 2003]. Bacterial amylase, however, is generally preferred over fungal amylase due to several characteristic advantages that it offers. Strains of *Aspergillus* sp. and *Bacillus* sp., mainly *Bacillus subtilis*, *B. stearothermophilus*, *B. amyloliquefaciens* and *B. licheniformis*, are known to be good producers of α -amylase and these have been widely used for commercial production of the

enzyme for various applications [Vihinen and Mantasala 1989; Pandey et al. 2000]. Several *Bacillus* sp. and thermostable *Actinomycetes* including *Thermomonospora* and *Thermoactinomyces* are versatile producers of the α -amylases [Ben et al. 1999]. The genus *Bacillus* produces a large variety of extracellular enzymes of which *amylases* and *proteases* are of significant industrial importance. An extremely thermostable α -amylase is available from the mesophile *B. licheniformis* [Morgan et al. 1981]. Alkaliphilic *Bacillus* strains often produce enzymes active at alkaline pH, including alkaline α -amylase, *protease* and *carboxymethylcellulase* [Horikoshi 1996].

2. Literature Review:

2.1 Structural and functional characteristics

O-Glycoside hydrolases (EC3.2.1.-) are a wide spread group of enzymes that hydrolyze the glycosidic bond between two or more carbohydrates, or between a carbohydrate and a non-carbohydrate moiety. A classification system for glycoside hydrolases, based on sequence similarity, has led to the definition of 85 different families. Most of the starch hydrolyzing enzymes belong to the α -amylase family or family 13 glycoside hydrolases based on amino acid sequence homology according to the classification of Henrissat (1991). The α -amylase family of glycoside hydrolases, is the largest family of glycoside hydrolases, transferases and isomerases comprising nearly 30 different enzyme specificities [Henrissat 1991]. A large variety of enzymes are able to act on starch. These enzymes are listed in Table 1.

These enzymes can be divide basically into four groups: endoamylases, exoamylases, debranching enzymes and transferases [M.J.E.C. van der Maarel et al. 2002]:

1. Endoamylases: cleave internal α -1,4 bonds resulting in α -anomeric products,
2. Exoamylases: cleave α -1,4 or α -1,6 bonds of the external glucose residues resulting in α - or β -anomeric products,
3. Debranching enzymes: hydrolyze α -1,6 bonds exclusively leaving long linear polysaccharides, and

4. Transferases: cleave α -1,4 glycosidic bond of the donor molecule and transfer part of the donor to a glycosidic acceptor forming a new glycosidic bond.

Table 1. Known activities of Glycoside hydrolase family 13 enzymes

Enzyme	EC number	Main substrate
Amylosucrase	EC: 2.4.1.4	Sucrose
Sucrose phosphorylase	EC: 2.4.1.7	Sucrose
Glucan branching enzyme	EC: 2.4.1.18	Starch, glycogen
Cyclomaltodextrin glycosyltransferase	EC: 2.4.1.19	Starch
Amylomaltase	EC: 2.4.1.25	Starch, glycogen
Maltopentaose-forming alpha-amylase	EC: 3.2.1.-	Starch
Alpha-amylase	EC: 3.2.1.1	Starch
Oligo-1,6-glucosidase	EC: 3.2.1.10	1,6-alpha-D-glucosidic linkages in some oligosaccharides
Alpha-glucosidase	EC: 3.2.1.20	Starch
Amylopullulanase	EC: 3.2.1.41	Pullulan
Cyclomaltodextrinase	EC: 3.2.1.54	linear and cyclomaltodextrin
Isopullulanase	EC: 3.2.1.57	Pullulan
Isoamylase	EC: 3.2.1.68	Amylopectin
Maltotetraose-forming alpha-amylase	EC: 3.2.1.60	Starch
Glucodextranase	EC: 3.2.1.70	Starch
Trehalose-6-phosphate hydrolase	EC: 3.2.1.93	Trehalose
Maltohexaose-forming alpha-amylase	EC: 3.2.1.98	Starch
Maltogenic amylase	EC: 3.2.1.133	Starch
Neopullulanase	EC: 3.2.1.135	Pullulan
Malto-oligosyl trehalase hydrolase	EC: 3.2.1.141	Trehalose
Malto-oligosyl trehalose synthase	EC: 5.4.99.15	Maltose

The α -glycosidic bond is very stable having a spontaneous rate of hydrolysis at room temperature [Wolfenden et al. 1998]. The catalytic mechanism of the α -amylase family is that of the α -

retaining double displacement [M.J.E.C. van der Maarel et al. 2002]. α -Retaining mechanism is the characteristic feature of the enzymes from the α -amylase family. They vary widely in their reaction specificities. The attachments of different domains to the catalytic site or to extra sugar binding subsites around the catalytic site is the prime reason for these differences [M.J.E.C. van der Maarel et al. 2002].

2.2 Starch

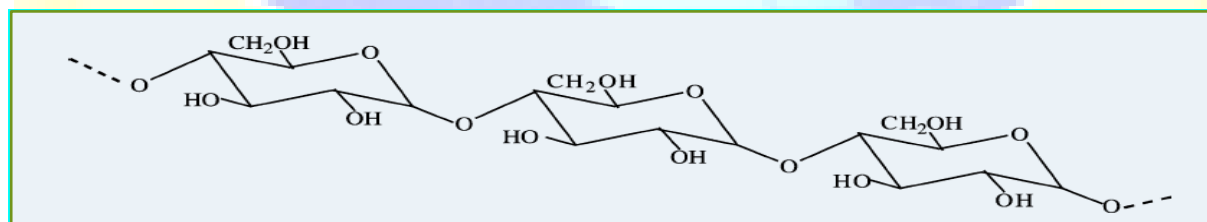
In the green leaves of plants carbon dioxide and water are transformed into glucose and oxygen under the influence of sunlight and with the help of chlorophyll. This process is known as photosynthesis. During the day this starch is deposited as grains in the leaf, the so-called leaf-transition starch. During the night this starch is partially broken down again into sugars which are transported to other areas of the plant. From these sugars the starch arises which is won in the familiar grain shape. The forming of starch is a process which has by far not been clarified yet and during which a number of enzymes play a role.

Starch or amyllum is a carbohydrate consisting of a large number of glucose units joined by glycosidic bonds. The major industrial sources are maize, tapioca, potato, and wheat, but limitations such as low shear resistance, thermal resistance, thermal decomposition and high tendency towards retrogradation limit its use in some industrial food applications [Goyal et al. 2005; M.J.E.C. van der Maarel et al. 2002]. With the help of a microscope the grain shape reveals from which plant species the starch derives. Native starch, the starch as it occurs in the plant, can not be dissolved in cold water. When we scatter starch, while stirring, into water we get a milky white suspension which can be stirred without much difficulty. When the stirring is stopped the starch sinks to the bottom (sedimentation), during which process a transparent upper layer is formed. When the suspension is heated the white colour disappears at a temperature characteristic for starch. The starch dissolves into an almost transparent solution. This is what we call gelatinized starch. In comparison with the ungelatinized suspension, stirring takes considerably more difficulty. The temperature at which the resistance during stirring noticeably increases, is called the gelatinization temperature. Gelatinizing starch into viscous substances (swellings) is one of the most, if not the most, important characteristic(s) of starch. This phenomenon lies at the basis of the successful application of starch in a large number of sectors.

Among carbohydrate polymers, starch is currently enjoying increased attention due to its usefulness in different food products. Starch contributes greatly to the textural properties of many foods and is widely used in food and industrial applications as a thickener, colloidal stabilizer, gelling agent, bulking agent and water retention agent [Jaspreet Singh et al. 2007]. Starch is a polymer of glucose linked to another one through the glycosidic bond. Two types of glucose polymers are present in starch: amylose and amylopectin (Fig.1). Amylose and amylopectin have different structures and properties. Amylose is a linear polymer consisting of up to 6000 glucose units with α -1,4 glycosidic bonds. Amylopectin consists of short α -1,4 linked to linear chains of 10–60 glucose units and α -1,6 linked to side chains with 15–45 glucose units. Granule bound starch synthase can elongate maltooligosaccharides to form amylose and is considered to be responsible for the synthesis of this polymer. Soluble starch synthase is considered to be responsible for the synthesis of unit chains of amylopectin.

α -Amylase is able to cleave α -1,4 glycosidic bonds present in the inner part of the amylose or amylopectin chain [Muralikrishna and Nirmala 2005; Tester et al. 2004; M.J.E.C. van der Maarel et al. 2002].

A. Structure of amylose



B. Structure of amylopectin

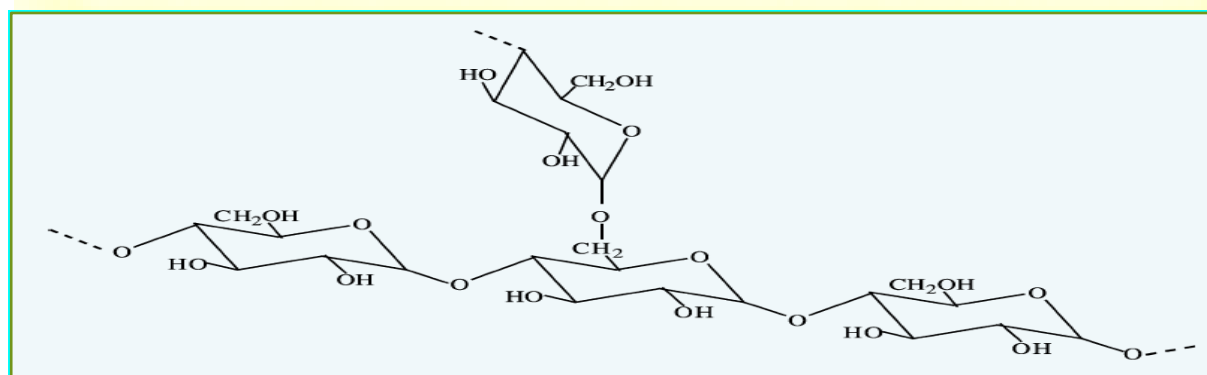


Figure 1. Types of glucose polymers present in starch: amylose (A) and amylopectin (B). From Muralikrishna and Nirmala (2005).

Endoamylases are able to cleave α ,1-4 glycosidic bonds present in the inner part (endo-) of the amylose or amylopectin chain. α -Amylase (EC3.2.1.1) is a well-known *endoamylase*. It is found in a wide variety of microorganisms [Pandey et al. 2000]. The end products of α -amylase action are oligosaccharides with varying length with an α -configuration and α -limit dextrins, which constitute branched oligosaccharides, which is one of the most important commercial enzyme processes. Saccharide composition obtained after hydrolysis of starch is highly dependent on the effect of temperature, the conditions of hydrolysis and the origin of enzyme. Specificity, thermostability and pH response of the enzymes are critical properties for industrial use [Kandra 2003].

α -Amylase finds application in all the industrial processes such as in food, detergents, textiles and in paper industry, for the hydrolysis of starch [Gupta et al. 2003; Konsula and Liakopoulou-Kyriakides 2004; Tanyildizi et al. 2005]. *Exoamylases* act on the external glucose residues of amylose or amylopectin and thus produce only glucose (*glucoamylase* and α -*glucosidase*), or maltose and β -limit dextrin (β -*amylase*).

A number of reviews exist on amylases and their applications, however, none specifically covers α -amylases at length. α -Amylases are one of the most popular and important form of industrial amylases and the present review highlights the various aspects of microbial α -amylases.

2.3 Microbial α -amylase production

Commercial sources of enzymes are obtained from three primary sources, i.e., animal tissue, plants and microbes. These naturally occurring enzymes are quite often not readily available in sufficient quantities for food applications or industrial use. However, by isolating microbial strains that produce the desired enzyme and optimizing the conditions for growth, commercial quantities can be obtained. This technique, well known for more than 3,000 years, is called fermentation. The enzymes are inducible, i.e., produced only when needed, and they contribute to the natural carbon cycle. Several methods, such as submerged fermentation (SmF) and solid-state

fermentation (SSF) have been successfully used for α -amylase production from various microorganisms. Agro-industrial residues such as wheat bran, spent brewing grain, maize bran, rice bran, rice husk, coconut oil cake, mustard oil cake, corn bran, etc., are generally considered the best substrates for processes [Sodhi et al. 2005; Francis et al. 2003; Babu and Satyanarayana 1995; Baysal et al. 2003; Ramachandran et al. 2004; Vishwanathan and Surlikar 2001]. In addition, the utilization of these agroindustrial wastes, on one hand, provides alternative substrates and, on the other, helps in solving pollution problems, which otherwise may cause their disposal [Pandey et al. 1999].

Various agro-industrial residues (agrosubstrates) used for microbial α -amylase production are shown in Table 2.

Table 2. Various agro-industrial residues (agrosubstrates) used for α -amylase production

Substrate	Organism	Activity, U/g	Reference
Wheat bran	<i>Bacillus</i> sp. PS-7	464 000	[Sodhi et al. 2005]
Spent brewing grain	<i>A. oryzae</i> NRRL 6270	6 583	[Francis et al. 2003]
Maize bran	<i>B. coagulans</i>	22 956	[Babu and Satyanarayana 1995]
Rice bran	<i>Bacillus</i> sp. PS-7	145 000	[Sodhi et al. 2005]
Rice husk	<i>B. subtilis</i>	21 760	[Baysal et al. 2003]
Coconut oil cake	<i>A. oryzae</i>	3 388	[Ramachandran et al. 2004]
Mustard oil cake	<i>B. coagulans</i>	5 953	[Babu and Satyanarayana 1995]
Corn bran	<i>Bacillus</i> sp. PS-7	97 600	[Sodhi et al. 2005]
<i>Amaranthus</i> grains	<i>Aspergillus flavus</i>	1 920	[Vishwanathan and Surlikar 2001]
Gram bran	<i>B. coagulans</i>	8 984	[Babu and Satyanarayana 1995]

α -Amylase may be derived from several bacteria, yeasts and fungi, but for commercial applications α -amylase is mainly derived from the genus *Bacillus*. Bacterial amylase, however, is generally

preferred over fungal *amylase* due to several characteristic advantages that it offers. Strains of *Aspergillus sp.* and *Bacillus sp.*, mainly *Bacillus amyloliquefaciens* and *B. licheniformis*, are employed for commercial applications. Thermostable α -amylases

are generally preferred as their application minimizes contamination risk and reduces reaction time, thus providing considerable energy saving. Hydrolysis carried out at higher temperatures also minimizes polymerization of D-glucose to iso-maltose.

Most reports about fungi that produce α -amylase have been limited to a few species of mesophilic fungi, mostly to *Aspergillus* and *Penicillium* [Gupta et al. 2003; Kathiresan and Manivannan 2006]. Filamentous fungi, such as *Aspergillus oryzae* and *Aspergillus niger*, produce considerable quantities of enzymes that are used extensively in the industry [Djekrif-Dakhmouche et al. 2005; Hernandez et al. 2006; Jin et al. 1998; Kammoun et al. 2008]. The thermophilic fungus *Thermomyces lanuginosus* is an excellent producer of *amylase* [Jensen et al. 2002; Kunamneni et al. 2005].

The fungal α -amylases are preferred over other microbial sources due to their more accepted GRAS (Generally Recognized As Safe) status [Gupta et al. 2003].

Optimization of various parameters and manipulation of media are one of the most important techniques used for the overproduction of enzymes in large quantities to meet industrial demands [Tanyildizi et al. 2005]. Growth of mycelium is crucial for extracellular enzymes like α -amylase [Carlsen et al. 1996]. Various physical and chemical factors have been known to affect the production of α -amylase such as temperature, pH, period of incubation, carbon sources acting as inducers, surfactants, nitrogen sources, phosphate, different metal ions, moisture and agitation with regards to SSF and SmF, respectively.

There is a very huge demand to improve the stability of the enzymes to meet the requirements set by specific applications, especially with respect to temperature and pH.

The optimum temperature depends on whether the culture is mesophilic or thermophilic. Among the fungi, most *amylase* production studies have been done with mesophilic fungi within the temperature range of 25-37 °C [Ramachandran et al. 2004; Francis et al. 2003]. The temperature optimum for the activity of α -amylase is related to the growth of the microorganism [Vihinen et al. 1989]. Thermostabilities are affected by many factors like

presence of calcium, substrate and other stabilizers[Vihinen et al. 1989].

pH is one of the important factors that determine the growth of microorganisms as they are sensitive to the concentration of hydrogen ions present in the medium. α -Amylases are generally stable over a wide range of pH from 4 to 11 [Fogarty et al. 1979; Vihinen et al. 1989; Hamilton et al. 1999; Khoo et al. 1994], however, α -amylases with stability in a narrow range have also been reported [Coronado et al. 2000; Fogarty 1983]. Fungi of *Aspergillus* sp. were found to give significant yields of α -amylase at pH=5.0-6.0 in SmF [Hayashida and Teramoto 1986; Carlsen et al. 1996; Djekrif-Dakhmouche et al. 2005]. Bacterial cultures *Bacillus* sp. required an initial pH of 7.0 [Tanyildizi et al. 2005; Syu and Chen 1997; Haq et al. 2005].

Thermophilic anaerobic bacteria *Clostridium thermosulfurogenes* gave maximum titres of α -amylase at pH=7.0 [Swamy and Seenayya 1996]. In fungal processes, the buffering capacity of some media constituents sometimes eliminates the need for pH control [Chahal 1983]. The pH values also serve as a valuable indicator of the initiation and end of enzyme synthesis [Friedrich et al. 1989]. It is reported that *A. oryzae* 557 accumulated α -amylase in the mycelia when grown in phosphate or sulphate deficient medium and was released when the mycelia were replaced in a medium with alkaline pH (above 7.2) [Yabuki et al. 1977].

Industrial enzymes produced in bulk generally require little downstream processing and hence are relatively crude preparations. The commercial use of α -amylase generally does not require purification of the enzyme, but enzyme applications in pharmaceutical and clinical sectors require high purity amylases. Laboratory scale purification for α -amylase includes various combinations of ion exchange, gel filtration, hydrophobicity interactions and reverse phase chromatography. Alternatively, α -amylase extraction protocols using organic solvents such as ethanol, acetone and ammonium sulfate precipitation [Glymph and Stutzenberger 1977; Hamilton et al. 1999; Khoo et al. 1994] and ultrafiltration have been proposed [Moraes et al. 1999]. These conventional multi-step methods require expensive equipments at each step, making them laborious, time consuming, barely reproducible and may result in increasing loss of the desired product [Arauz et al. 2009].

Purification processes in downstream processing after fermentation strongly depend on the market, processing cost, final quality, and available technology. Most enzymes are purified by chromatographic techniques after crude isolation by precipitation and membrane separations. The need for large-scale cost effective purification of proteins has resulted in evolution of techniques

that provide fast, efficient and economical protocols in fewer processing steps [Amritkar et al. 2004]. Purification techniques that produce homogeneous preparation of α -amylases in a single step are given in Table 3.

Table 3. Methods of one-step purification of α -amylases

Method	Adsorbent	Yield/ %	Purification fold	Reference
Affinity adsorption chromatography	β -cyclodextrin- iminodiacetic acid- Cu^{2+}	95	–	[Liao and Syu 2005]
Expanded bed chromatography	Alginate acid-cellulose cell beads	69	51	[Amritkar et al. 2004]
High speed counter current chromatography	PEG4000-aqueous two-phase system	73.1	–	[Zhi et al. 2005]
Magnetic affinity adsorption	Magnetic alginate microparticles	88	9	[Safarikova et al. 2003]
Substitute affinity method	Insoluble corn starch at 4 °C	78	163	[Najafi and Kembhavi 2005]

2.4 Industrial applications of α -amylase

The history of the industrial production of enzymes dates back to the time when Dr. Jokichi Takamine began the production of digestive enzyme preparation of α -amylase from *A. oryzae* in 1894 known as »Taka diastase«, which was used as a digestive aid.

Amylases are among the most important hydrolytic enzymes for all starch based industries, and the commercialisation of amylases is oldest with first use in 1984, as a pharmaceutical aid for the treatment of digestive disorders.

Such enzymes hydrolyze the starch molecules into polymers composed of glucose units. Amylases have found applications in starch processing, desizing of textiles, paper sizing, as

detergent additive, and bread improvement, pharmaceutical industries, ethanol, and other fermentation processes [Haki and Rakshit 2003; Lowe 2002; Gomes et al. 2005].

The global market for enzymes was about \$2 billion in 2004. It is expected to have an average annual growth rate of 3.3 %. The share of carbohydrases comprising *amylases*, *isomerases*, *pectinases* and *cellulases* is about 40 % [Riegal and Bissinger 2003]. The food and beverage sectors utilize 90 % of the carbohydrases produced. Today, *amylases* have the major world market share of enzymes [Aehle and Misset 1999]. In Table 4 are shown applications of *amylases* in various sectors of industry. In this light, microbial *amylases* have completely replaced chemical hydrolysis in the starch processing industry.

Table 4. Uses of amylases in various sectors of industry

Sector	Uses	Reference
Food industry	Production of glucose syrups, crystalline glucose; Production of high fructose corn syrups; Production of maltose syrups; Reduction of viscosity of sugar syrups; Reduction of haze formation in juices; Solubilization and saccharification of starch for alcohol fermentation in brewing industries; Retardation of staling in baking industry;	-[M.J.E.C. van der Maarel et al. 2002] -[Riegal and Bissinger 2003] -[Gupta et al. 2003] - [Haki and Rakshit 2003] - [Lowe 2002] -[Gomes et al. 2005].
Detergent industry	Used as an additive to remove starch based dirt	
Paper industry	Reduction of viscosity of starch for appropriate coating of paper	
Textile industry	Warp sizing of textile fibers	
Pharmaceutical industry	Used as a digestive aid	

A comprehensive account on commercial applications of α -amylases is quoted by Godfrey and West (1996). Various applications of α -amylase are dealt here in brief.

2.4.1 Food industry

Amylases are extensively employed in processed-food industry such as baking, brewing, preparation of digestive aids, production of cakes, fruit juices and starch syrups [Couto and Sanromán 2006]. For decades, microbial α -amylases have been widely used in the baking industry [Hamer 1995; Si 1999]. These enzymes can be added to the dough of bread to degrade the starch in the flour into smaller dextrins, which are subsequently fermented by the yeast. Besides generating fermentable compounds, α -amylases also have an anti-staling effect in bread baking, and they improve the softness retention of baked goods, increasing the shelf life of these products [Gupta et al. 2003; M.J.E.C. van der Maarel et al. 2002; Sahlstrom and Brathen 1997]. The addition of α -amylase to the dough results in enhancing the rate of fermentation and the reduction of the viscosity of dough, resulting in improvements in the volume and texture of the product.

The most widespread applications of α -amylases are in the starch industry, which are used for starch hydrolysis in the starch liquefaction process that converts starch into fructose and glucose syrups [Nielsen and Borchert 2000; Gupta 2003]. The hydrolysis of starch may be carried out using either acid or enzyme as catalyst. Acid conversion has, however, many limitations: it is non-specific, lacks ways of controlling saccharide composition, requires high refining costs and is less environmentally friendly. The application of enzymes for this process has avoided these limitations [Crabb and Shetty 1999]. Conversion of starch into sugar, syrups and dextrins forms the major part of the starch processing industry. *Amylases* are also used for the clarification of beer or fruit juices, or for the pretreatment of animal feed to improve the digestibility of fiber [Gavrilescu and Chisti 2005; Ghorai et al. 2009; M.J.E.C. van der Maarel et al. 2002].

2.4.2 Textile industry

Amylases are used in textile industry for desizing process. Sizing agents like starch are applied to yarn before fabric production to ensure a fast and secure weaving process. In textile weaving, starch paste is applied for warping. This gives strength to the textile at weaving. It also prevents the

loss of string by friction, cutting and generation of static electricity on the string by giving softness to the surface of string due to laid down warp.

After weaving the cloth, the starch is removed and the cloth goes to scouring and dyeing. The starch on cloth is usually removed by application of α -amylase [Hendriksen et al. 1999].

Starch is a very attractive size, because it is cheap, easily available in most regions of the world, and it can be removed quite easily. Starch is later removed from the woven fabric in a wet-process in the textile finishing industry.

The enzymatic desizing of cotton with α -amylases is state-of-the-art since many decades [Marcher et al. 1993]. The amylose is bioconverted to 100% by the α -amylase into glucose whereas the amylopectin is converted to 50% into glucose and maltose. Bio-desizing is preferred due to their high efficiency and specific action. Amylases bring about complete removal of the size without any harmful effects on the fabric besides eco friendly behavior.

The α -amylases remove selectively the size and do not attack the fibres [Ahlawat et al. 2009; Feitkenhauer 2003; Gupta 2003]. Amylase from *Bacillus* stain was employed in textile industries for quite a long time.

2.4.3 Paper industry

The use of α -amylase in pulp and paper industry is in the modification of starches for coated paper, i.e. for the production of low-viscosity, high molecular weight starch. As for textiles, sizing of paper with starch is performed to protect the paper against mechanical damage during processing [Gupta et al. 2003; M.J.E.C. van der Maarel et al. 2002; Bruinenberg et al. 1996]. The coating treatment serves to improve the quality of the finished product, enhances stiffness, and elasticity of paper [Gupta et al. 2003; Bruinenberg et al. 1996]. Because starch is added to paper at a temperature range of 45- 60 °C, and the viscosity of the natural starch is too high for paper sizing partial degradation of this polymer is essential. α -Amylase is employed for this purpose [Gupta et al. 2003].

2.4.4 Detergent applications

The demand for α -amylase for use in laundry and automatic dishwashing is very high. The use of enzymes in detergent formulations enhances the detergent's ability to remove tough stains and making the detergent environmentally safe. These enzymes are used in detergents for laundry and automatic dishwashing to degrade the residues of starchy foods such as potatoes, gravies, custard, chocolate, etc. to dextrins and other smaller oligosaccharides [Mukherjee et al. 2009; [Olsen and Falholt 1998]. Removal of starch from surfaces is also

important in providing a whiteness benefit, since starch can be an attractant for many types of particulate soils. 90% of all liquid detergents contain these enzymes [Gupta et al. 2003].

The oxidative stability of amylases is one of the most important criteria for their use in detergents where the washing environment is very oxidizing [Kirk et al. 2002].

Alkaliphilic *Bacillus* strains often produce enzymes active at alkaline pH, including alkaline α -amylase [Horikoshi, 1996]. When alkaline α -amylase is used as a component of detergents, the chelating agents usually contained in detergents easily remove calcium, which is essential for its stability. Thus there is a search for Ca free α -amylase [Nonaka et al., 2003].

2.4.5 Beverage alcohol and Fuel Ethanol production

In beer industries microbial amylases are used to aid cereal amylase in the production of fermentable sugar. Ethanol is the most utilized liquid biofuel. Over the past decades, there has been an increasing interest in fuel ethanol as a result of increased environmental concern and higher crude oil prices. Ethanol fuels can be derived from renewable resources such

as agricultural crops and by products. For the ethanol production, starch is the most used substrate due to its low price and easily available raw material in most regions of the world [Chi et al. 2009].

The bioconversion of starch into ethanol involves liquefaction and saccharification, where starch is converted into sugar using an amylolytic microorganism or enzymes such as α -amylase, followed by fermentation, where sugar is converted into ethanol using an ethanol fermenting microorganism such as yeast *Saccharomyces cerevisiae* [Moraes et al. 1999].

Enzymes such as *α -amylase*, *glucoamylase* and *cellulases* are important to produce fermentable sugars to produce ethanol [Kirk et al. 2002].

2.4.6 Treatment of starch processing waste water

Starch is also present in waste produced from food processing plants. Starch waste causes pollution problems. Biotechnological treatment of food processing waste water can produce valuable products such as microbial biomass protein and also purifies the effluent [Friedrich et al. 1987; Kingspohn et al., 1993].

2.4.7 Other applications

The spectrum of amylase application has widened in many other fields, such as clinical, medical, and analytical chemistries [Pandey et al., 2000; Cherry et al., 2004]. To some extent *amylases* are also used as digestive aids to supplement the diastatic activity of flour and to improve digestibility of some of the animal feed ingredients [Kumar et al. 1995].

There are several processes in the medicinal and clinical areas that involve the application of *amylases*. The application of a liquid stable reagent, based on *α -amylase* for the Ciba Corning Express clinical chemistry system has been described [Becks et al. 1995]. A process for the detection of higher oligosaccharides, which involved the application of *amylase* was also developed [Giri et al. 1990].

3. Future Directions:

As evident from the foregoing review, *amylases* are among the most important enzymes used in industrial processes. Although, the use of *α -amylase* in starch based industries has been prevalent for many decades and a number of microbial sources exist for the efficient production of this enzyme, but only a few selected strains of fungi and bacteria meet the criteria for commercial production.

The continued development of new enzymes through modern biotechnology may, for example, lead to enzyme products with improved cleaning effects at low temperatures. This could allow wash temperatures to be reduced, saving energy in countries where hot washes are still used.

Today, white biotechnology is geared towards creating new materials and biobased fuels from agricultural waste and providing alternative biobased routes to chemical processes. These efforts could lead to the development of improved enzymes such as *amylases*, *hemicellulases* or *cellulases* that could be used in the industries.

Today, the application of biotechnology to industrial processes holds many promises for sustainable development. New and exciting enzyme applications are likely to bring benefits in other areas: less harm to the environment; greater efficiency; lower costs; lower energy consumption; and the enhancement of a product's properties.

The use of enzymes not only make the process less toxic (by substituting enzymatic treatments for harmful chemical treatments) and eco-friendly, they reduce costs associated with the production process, and consumption of natural resources (water, electricity, fuels), while also improving the quality of the final product. It seems that in the future it will be possible to do every process using enzymes.

4. Conclusion:

Amylases are important in many industrial processes and are one of the most widely used enzymes required for the preparation of fermented foods. Apart from food and starch industries, in which demand for them is increasing continuously, they are also used in various other industries such as paper and pulp, textile, etc. To alleviate the ever-growing demand for fuel energy the production of fuel ethanol from plant material is the focus of research. To this effect the application of amylolytic enzymes in the production of the fermentable sugar from starchy crops is indispensable. Starch degrading enzymes like *amylase* have received great deal of attention because of their perceived technological significance and economic benefits.

Pollution free processes are gaining ground all over the world. Enzymes are not only beneficial from ecological point of view but they are also saving lot of money by reducing water and energy consumption which ultimately reduce the cost of production.

A number of microbial sources exist for the efficient production of this enzyme, but only a few selected strains of fungi and bacteria meet the criteria for commercial production. In order to achieve the efficient, large-scale production, the structural and functional relationships of α -

amylases have to be known in detail. This will lead to improving the stability of the existing enzymes and discovery of many new ones.

5. REFERENCES:

- Aehle, W., Misset, O., (1999). Enzymes for industrial applications. In: Rehm HJ, Reed G, editors. *Biotechnology*, 2nd ed. Germany: Wiley-VCH, 189-216.
- Ahlawat, S., Dhiman, S.S., Battan, B., Mandhan, R.P., Sharma, J., (2009). Pectinase production by *Bacillus subtilis* and its potential application in biopreparation of cotton and micropoly fabric. *Process Biochemistry*, 44, 521–526.
- Amritkar, N., Kamat, M., Lali, A., (2004). Expanded bed affinity purification of bacterial α -amylase and cellulase on composite substrate analogue–cellulose matrices, *Process Biochem.*, 39, 565–570.
- Arauza, L.J., Jozalaa, A.F., Mazzolab, P.G., Penna, T.C.V., (2009). Nisin biotechnological production and application: a review. *Trends Food Sci Technol.*, 20, 146-154.
- Babu, K.R., Satyanarayana, T., (1995). α -Amylase production by thermophilic *Bacillus coagulans* in solid-state fermentation, *Process Biochem.*, 30, 305–309.
- Baysal, Z., Uyar, F., Aytekin, C., (2003). Solid-state fermentation for production of α -amylase by a thermotolerant *Bacillus subtilis* from hot-spring water, *Process Biochem.*, 38, 1665–1668.
- Becks, S., Bielawski, C., Henton, D., Padala, R., Burrows, K., Slaby, R., (1995). Application of a liquid stable amylase reagent on the CibaCorning Express clinical chemistry system. *Clin Chem.*, 41, 186.
- Ben, Am., Mezghani, M., Bejar, S., (1999). A thermostable α -amylase producing maltohexaose from a new isolated *Bacillus* sp. US100: study of activity and molecular cloning of the corresponding agent. *Enzyme. Microb. Technol.*, 24, 548-549.
- Bordbar, A.K., Omidian, K., Hosseinzadeh, R., (2005). Study on interaction of α -amylase from *Bacillus subtilis* with cetyltrimethylammonium bromide, *Colloids surf. B: Biointerfaces*, 40, 67-71.
- Bruinenberg, P.M., Hulst, A.C, Faber, A., Voogd, R.H., (1996). A process for surface sizing or coating of paper. European Patent Application EP 0,690,170 A1.

- Burhan, A., Nisa, U., Gokhan, C., Omer, C., Ashabil, A., Osman, G., (2003). Enzymatic properties of a novel thermostable, thermophilic, alkaline and chelator resistant *amylase* from an alkaliphilic *Bacillus* sp. Isolate ANT-6, *Process. Biochem.*, 38, 1397-1403.
- Carlsen, A.B. Spohr, J. Nielsen, J. Villadsen, (1996). Morphology and physiology of an α -amylase producing strain of *Aspergillus oryzae* during batch cultivations, *Biotechnol. Bioeng.*, 49, 266–276.
- Cavaco-Paulo A. & Gübitz G. M., (2003). Cambridge: Woodhead Publishing, *Textileprocessing with enzyme*. 17–18, 30–34, 51–52, 90–95, 110, 124–125, 129–131, 158–169, ISBN 18557366101.
- Chahal, D.S., (1983). Growth characteristics of microorganisms in solidstate fermentation for upgrading of protein values of lignocellulosesand cellulase production. In: Blanch HW, Papoutsakis ET, Stephanopoulos G, editors. Foundations of biochemical engineering kinetics and thermodynamics in biological systems. American Chemical Society, Washington DC. ACS symposium series, No. 207, 1983, 421-442.
- Chelly, M., Dahan, R., Labyod, M., Dupont, F., Lacoste, C. and Larue, F. (1996). *Ann. Pediatr.* 43, 392–402.
- Cherry, H.M., Hossain, M. T., and Anwar ,M..N. (2004). Extracellular Glucoamylase from the Isolate *Aspergillus fumigatus*. *Pakit J Biol. Sci*, 7 (11), 1988-1992.
- Chi, Z., Chi, Z., Liu, G., Wang, F., Ju, L., Zhang, T., (2009). *Saccharomycopsis fibuligera* and its applications in biotechnology. *Biotechnol Adv.*, 27, 423-431.
- Chiu, L. Y. and Chandler, W. L., (1995). *Clin. Chem.*, 41, 238–239.
- Ciechańska D and Kazimierzak J., (2006). *Fibres 4. & Textiles in Eastern Europe*, 14, No 1(55), 92-95.
- Coronado, M., Vargas, C., Hofemeister, J., Ventosa, A., Nieto, J.J., (2000). Production and biochemical characterization of an α -amylase from the moderate halophile *Halomonas meridiana*. *FEMS Microbiol. Lett.*, 183, 67-71.
- Couto, S.R., Sanromán, M.A., (2006). Application of solid-state fermentation to food industry- A review. *Journal of Food Engineering*, 76, 291-302.
- Crueger, W. and Crueger, A., (1989). Industrial Microbiology (eds.), Sinauer Associates, Sunderland, MA, 189–218.

- Djekrif-Dakhmouche, S.; Gheribi-Aoulmi, Z.; Meraihi, Z.; Bennamoun, L. (2005). Application of a statistical design to the optimization of culture medium for α -amylase production by *Aspergillusniger* ATCC 16404 grown on orange waste powder. *J Food Process Eng* 73, 190–197.
- Feitkenhauer, H. (2003). Anaerobic digestion of desizing wastewater: influence of pretreatment and anionic surfactant on degradation and intermediate accumulation. *Enzyme Microb. Technol.*, 33, 250–258.
- Fogarty, M.W., Kelly, C.T. (1979). Developments in microbial extracellular enzymes. In: Wiseman A, editor. *Topics in enzyme and fermentation biotechnology*, 3, 45–108.
- Fogarty, M.W., Kelly, C. T., (1980). In *Economic Microbiology, Microbial Enzymes and Bioconversions*, vol. 5 (Rose, A. H., ed.), Academic Press, London, 115–170.
- Fogarty, M.W., (1983). Microbial Amylases. In: *Microbial Enzymes and Biotechnology*, Fogarty, M.W. (Ed.), Applied Science Publishers Ltd., London, UK, 1–92.
- Francis, F., Sabu, A., Nampoothiri, K.M., Ramachandran, S., Ghosh, S., Szakacs, G., Pandey, A., (2003). Use of response surface methodology for optimizing process parameters for the production of α -amylase by *Aspergillusoryzae*, *Biochem. Eng. J.*, 15, 107–115.
- Friedrich, J., Cimerman, A., Perdih, A., (1987). Mixed culture of *Aspergillus awamori* and *Trechoderma reesi* for bioconversion of apple distillery waste. *Appl. Microbiol. Biotechnol.*, 26, 299–305.
- Friedrich, J., Cimerman, A., Steiner, W., (1989). Submerged production of pectinolytic enzymes by *Aspergillus niger* : effect of different aeration/agitation regimes. *Appl Microbiol Biotechnol*, 31, 490–494.
- Gavrilescu, M., Chisti, Y., (2005). Biotechnology-a sustainable alternative for chemical industry. *Biotechnol Adv*, 23, 471–499.
- Ghorai, S., Banik, S.P., Verma, D., Chowdhury, S., Mukherjee, S., Khowala, S., (2009). Fungal biotechnology in food and feed processing. *Food Res. Int.*, 42, 577–587.
- Giri, N.Y., Mohan, A.R., Rao, L.V., Rao, C.P., (1990). Immobilization of α -amylase complex in detection of higher oligosaccharides on paper. *Curr Sci.*, 59, 1339–1340.
- Glymph, J.L., Stutzenberger, F.J., (1977). Production, purification, and characterization of α -amylase from *Thermomonospora curvata*. *Appl Environ Microbiol*, 34, 391–397.

- Godfrey, T., West, S., (1996). In: Godfrey, T., West, S., editors. *Industrialenzymology*. 2nd ed. New York: Stockton Press, 91,105-131,192,339-356,361-371.
- Gomes, E., Sauza, S. R., Grandi,R. P.and Silva, R. (2005). Glucoamylasesproduced from *A. flavus* A1.1 and *T. Braz. J. Microbiol.*,36(1), 75-82.
- Goyal, N., Gupta, J.K.,Soni, S.K., (2005). A novel raw starch digesting thermostable α -amylase from *Bacillus* sp. I-3 and its use in the direct hydrolysis of raw potato starch. *Enzyme Microb. Technol.* 37, 723–734.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V.K.,Chauhan, B., (2003). Microbial α -amylases: a biotechnological perspective. *Process Biochem*, 38, 1599 - 1616.
- Haki, G. D., Rakshit, S. K., (2003). Developments in industrially important thermostable enzymes: a review. *Bioresour Technol.*,89, 17-34.
- Hamer, R.J., (1995). Enzymes in the baking industry. In: Tucker, G.A.,Woods, L.F.J., editors. *Enzymes in food processing*. Galsgow:Blackie Academic and Professional, 190-222.
- Hamilton, L.M., Kelly, C.T., Fogarty, W.M., (1999). Purification and propertiesof the raw starch degrading α -amylase of *Bacillus* sp.IMD434.*Biotechol. Lett.*, 21,111-115.
- Haq, I., Ashraf, H., Qadeer, M.A., Iqbal, J., (2005). Pearl millet, asource of alpha amylase production by *Bacillus licheniformis*,*Bioresour. Technol.*,96,1201–1204.
- Hayashida, S., Teramoto, Y., (1986). Production and characteristicsof raw-starch-digesting α -amylase from a protease negative*Aspergillus ficuum* mutant, *Appl. Environ. Microbiol.*,52, 1068–1073.
- Hendriksen, H.V., Pedersen, S., Bisgard-Frantzen, H., (1999). A process for textile warp sizing using enzymatically modified starches. Patent Application WO 99/35325.
- Henrissat, B., (1991). A classification of glycosyl hydrolases based on amino acid sequence similiarities, *Biochem. J.*, 280, 309-316.
- Hernández, M.S., Rodríguez, M.R., Guerra, N.P., Rosés, R.P., (2006). Amylase production by *Aspergillusniger* in submerged cultivation on two wastes from food industries. *J Food Process Eng* 73, 93–100.
- Horikoshi, K., (1996). Alkaliphiles from an industrial point of view. *FEMS Microbiol. Rev.* 18, 259-270.

- JaspreetSingha, J., Kaurb, L., McCarthy, O.J., (2007). Factors influencing the physico-chemical, morphological, thermal and rheological properties of some chemically modified starches for food applications-A review. *Food Hydrocolloids*, 21, 1–22.
- Jenkins R. O. (2003). in: Textile Processing with Enzymes, Edited by Cavaco-Paulo A. & Gübitz GM, Woodhead publishing Ltd., CRC Press, Boca Raton, ISBN 18557366101.
- Jensen, B., Nebelung, P., Olsen, J., Reeslev, M., (2002). Enzyme production in continuous cultivation by the thermophilic fungus, *Thermomyces lanuginosus*. *Biotechnology Letters*, 24, 41–45.
- Jin, B., van Leeuwen, H.J., Patel, B., Yu, Q., (1998). Utilisation of starch processing wastewater for production of microbial biomass protein and fungal α -amylase by *Aspergillus oryzae*. *Bioresour. Technol.* 66, 201- 206.
- Kammoun, R., Naili, B., Bejar, S., (2008). Application of a statistical design to the optimization of parameters and culture medium for alpha-amylase production by *Aspergillus oryzae* CBS 819.72 grown on gruel (wheat grinding by-product). *Bioresour Technol*, 99, 5602-5609.
- Kandra, L., (2003). α -Amylases of medical and industrial importance. *Journal of Molecular Structure (Theochem)*, 666–667, 487–498.
- Kathiresan, K. and Manivannan, S. (2006). α -Amylase production by *Penicillium fellutanum* isolated from mangrove rhizosphere soil. *Afr. J. Biotechnol.* 5, 829-832.
- Khoo, S.L., Amirul, A.A., Kamaruzaman, M., Nazalan, N., Azizan, M.N., (1994). Purification and characterization of alpha-amylase from *Aspergillus flavus*. *Folia Microbiol (Praha)*, 39, 392-398.
- Kingspohn, K., Bader, J., Kruse, P.V., Schugerl, K., (1993). Utilization of potato pulp from potato starch processing. *Proc. Biochem.*, 28, 91-98.
- Konsula, Z., Liakopoulou-Kyriakides, M., (2004). Hydrolysis of starches by the action of an α -amylase from *Bacillus subtilis*. *Process Biochem*, 39, 1745–1749.
- Kumar, S. S., Venkateswara, R. M. and Das, D., (1990). Studies on glucoamylase produced from *Aspergillus awamori* (NRRL-3112) and their effect on saccharification of potato starch. *Indian J Exp Biol*, 33(12), 957-961.
- Lange, N.K. (1997). *Textile Chemist and Colorist*, 29, 23-26.

- Liao, Y.C., Syu, M.J., (2005). Novel immobilized metal ion affinity adsorbent based on cross-linked β -cyclodextrin matrix for repeated adsorption of α -amylase, *Biochem. Eng. J.*, 23, 17–24.
- Lowe, D.A., (2002). Production of enzymes In : *Basic biotechnology 2nd edn* (Colin Ratledge and Bjarn Kristiansen eds) Cambridge University press, UK, 391 – 407.
- Marcher, D., H.A. Hagen and S. Castelli, (1993). *ITB Veredlung*, 39, 20.
- Menzel, C., Lerch, T., Schneider, K., Weidemann, R., Tollnick, C., Kretzmer, G., Scheper, T. and Schugerl, K., (1998). *Process Biochem.* 33, 175–180.
- M.J.E.C. van der Maarel, B. van der Veen, Uitdehaag, J.C.M., Leemhuis, H., Dijkhuizen, L., (2002). Properties and applications of starch-converting enzymes of the α -amylase family, *J. Biotechnol.*, 94, 137-155.
- Moraes, L.M.P., Filho, S.A., Ulhoa, C.J., (1999). Purification and some properties of an α -amylase glucoamylase fusion protein from *Saccharomyces cerevisiae*. *World J. Microbiol. Biotechnol.*, 15, 561-564.
- Morgan, F.J., Priest, F.G., (1981). Characterization of a thermostable α -amylase from *Bacillus licheniformis* NCIB6346. *J. Appl. Bacteriol.* 50, 107-114.
- Mukherjee, A.K., Borah, M., Raí, S.K., (2009). To study the influence of different components of fermentable substrates on induction of extracellular α -amylase synthesis by *Bacillus subtilis* DM-03 in solid state fermentation and exploration of feasibility for inclusion of α -amylase in laundry detergent formulations. *Biochem. Eng. J.*, 43, 149–156.
- Muralikrishna, G., Nirmala, M., (2005). Cereal α -amylases – an overview. *Carbohydrate Polymers* 60, 163-173.
- Nielsen, J.E., Borchert, T.V., (2000). Protein engineering of bacterial α -amylases. *Biochim Biophys Acta*, 1543, 253-274.
- Nonaka, T., Fujihashi, M., Kita, A., Hagihara, H., Ozaki, K., Ltso, S. and Miki, K., (2003). Crystal structure of Calcium free α amylase from *Bacillus sp.* Strain KSM-K38 (amyK38) and its sodium ion Binding sites. *J. Biol. Chem.*, 278(27): 24828 – 24824.
- Olsen, H.S.O., Falholt, P., (1998). The Role of Enzymes in Modern Detergency. *Journal of Surfactants and Detergents*, 1, 555–567.
- Opwis, K., Knittel, D., Kele, A., Schollmeyer, E., (1999). *Starch/Starke*, 51, 348-353.

- Pandey, A., Nigam, P., Soccol, C.R., Soccol, V.Y., Singh, D. and Mohan, R., (2000). Advances in microbial amylases. *Biotechnol. Appl. Biochem.* 31, 135–152.
- Rajagopalan, G., Krishnan, C., (2008). Alpha-amylase production from catabolite repressed *Bacillus subtilis* KCC103 utilizing sugarcane bagasse hydrolysate. *Bioresour. Technol.*, 99, 3044–3050.
- Ramachandran, S., Patel, A.K., Nampoothiri, K.M., Francis, F., Nagy, V., Szakacs, G., Pandey, A., (2004). Coconut oil cake – A potential raw material for the production of α -amylase, *Bioresour. Technol.*, 93, 169–174.
- Reddy, N.S., Nimmagadda, A., Sambasiva Rao, K.R.S., (2003). An overview of the microbial α -amylase family. *Afr. J. Biotechnol.* 2, 645–648.
- Riegal, E.R., Bissinger, H.G., (2003). Industrial Fermentation: Principles, Processes and Products. In: *Riegal's Handbook of Industrial Chemistry*, J.A. Kent, (Ed.), Kluwer Academic/Plenum Publishers, New York, USA 963–1045.
- Safarikova, M., Roy, I., Gupta, M.N., Safarik, I., (2003). Magnetic alginate microparticles for purification of α -amylases, *J. Biotechnol.*, 105, 255–260.
- Sahlstrom, S., Brathen, E., (1997). Effects of enzyme preparations for baking, mixing time and resting time on bread quality and bread staling. *Food Chem.*, 58, 75–80.
- Si, J.Q., (1999). Enzymes, baking, bread making. In: Flickinger, M.C., Drew, S.W., editors. *Encyclopedia of bioprocess technology: fermentation, biocatalysis and bioseparation*, vol. 2. Wiley, 947–958.
- Sodhi, H.K., Sharma, K., Gupta, J.K., Soni, S.K., (2005). Production of a thermostable α -amylase from *Bacillus sp.* PS-7 by solid-state fermentation and its synergistic use in the hydrolysis of malt starch for alcohol production, *Process Biochem.*, 40, 525–534.
- Strandberg, A., Nystrom, A., Behr, S. and Karlsson, A. (1999). *Chromatography*, 50, 215–222.
- Sutton, A., Dawson, H., Hoff, B., Grift, E. and Shoukri, M., (1999). *Can. Vet. J.*, 40, 255–260.
- Swamy, M.V., Seenayya, G., (1996). Thermostable pullulanase and α -amylase activity from *Clostridium thermosulfurogenes* SV9 optimization of culture conditions for enzyme production, *Process Biochem.* 31, 157–162.

- Syu, M.J., Chen, Y.H., (1997). A study on the α -amylase fermentation performed by *Bacillus amyloliquefaciens*, *Chem. Eng. J.*, 65, 237–247.
- Tanyildizi, M.S., Ozer, D., Elibol, M., (2005). Optimization of α -amylase production by *Bacillus* sp. using response surface methodology, *Process Biochem*, 40, 2291–2296.
- Tavčer, P.F., (2011). Biotechnology in Textiles – an Opportunity of Saving Water, *Waste Water - Treatment and Reutilization*, ISBN: 978-953-307-249-4.
- Tester, R.F., Karkalas, J., Qi, X., (2004). Starch-composition, fine structure and architecture. *J. Cereal Sci.* 39, 151–165.
- Vihinen, M. and Mantasala, P., (1989). Microbial amylolytic enzymes, *Crit. Rev. Biochem. Mol. Biol.*, 24, 329-418.
- Vishwanathan, P., Surlikar, N.R., (2001). Production of α -amylase with *Aspergillus flavus* on Amaranthus grains by solid-state fermentation, *J. Basic Microbiol.*, 41, 57–64.
- Wolfenden, R., Lu, X., Young, G., (1998). Spontaneous hydrolysis of glycosides. *J. Am. Chem. Soc.*, 120, 6814-6815.
- Yabuki, M., Ono, N., Hoshino, K., Fukui, S., (1977). Rapid induction of α -amylase by non-growing mycelia of *Aspergillus oryzae*, *Appl Environ Microbiol*, 34, 1-6.
- Zhi, W., Deng, Q., Song, J., Gu, M., Ouyang, F., (2005). One-step purification of α -amylase from the cultivation supernatant of recombinant *Bacillus subtilis* by high-speed counter-current chromatography with aqueous polymer two-phase systems, *J. Chromatogr. A*, 1070, 215–219.